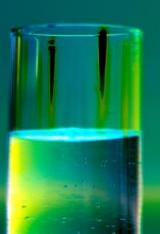
SSERC, 12 February 2014

Immunology Discussion Gary Entrican & Sean Wattegedera Moredun www.moredun.org.uk

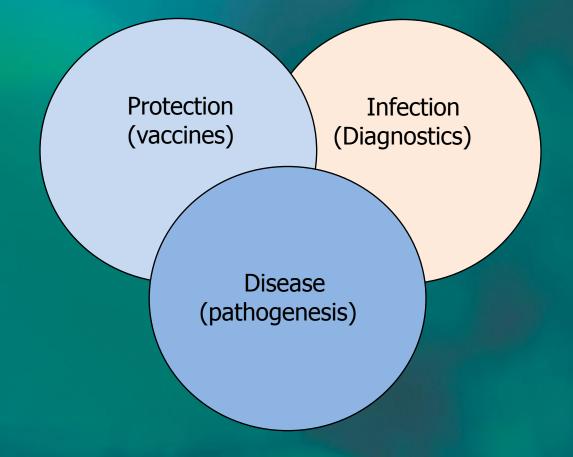








Uses of immunology





Curriculum elements relating to the immune system

Non-specific defences(i) Physical and chemical defences(ii) Inflammatory response(iii) Non-specific cellular responsesSpecific cellular defences(i) Immune surveillance(ii) Clonal selection theory(iii) T and B lymphocytes (Recognition of self and non self)(iv) Memory cells (immunological memory)

Infectious diseases and immunity

The transmission of infectious diseases Epidemiology of infectious diseases Active immunisation and vaccination Vaccine clinical trials Herd immunity Public health medicine Evasion of specific immune responses by pathogens: Antigenic variation Direct attack on the immune system



Higher Biology: Cell Biology Unit

Cellular response in defence in animals and plants

(i) The nature of viruses and their invasion of cells. Alteration of cell instructions to produce more viruses.

(ii) Cellular defense mechanisms in animals. Phagocytosis. Antibody production.

(iii) Cellular defence mechanisms in plants.

The importance of lysosomes in phagocytosis should be noted. Reference need not be made to different types of phagocytes. The production of antibodies by lymphocytes, and antibody action in response to the presence of foreign antigens, should be given simple treatment without reference to specific types of lymphocytes or to antigen-antibody reactions. The problem of tissue rejection and the use of suppressors in tissue transplantation.



Advanced Higher Biology (revised): Immune response to parasites

Non-specific defences of mammals: physical barriers, chemical secretions, inflammatory response, phagocytes and natural killer cells destroying abnormal cells.

Specific cellular defence in mammals involves immune surveillance by white blood cells, clonal selection of T lymphocytes, T lymphocytes targeting immune response and destroying infected cells by inducing apoptosis, phagocytes presenting antigens to B lymphocytes, the clonal selection of B lymphocytes, production of specific antibody by B lymphocyte clones, long term survival of some members of T and B lymphocyte clones to act as immunological memory cells.

Epidemiology is the study of the outbreak and spread of infectious disease. The herd immunity threshold is the density of resistant hosts in the population required to prevent an epidemic.

Endoparasites mimic host antigens to evade detection by the immune system, and modify host-immune response to reduce their chances of destruction. Antigenic variation in some parasites allows them to evolve fast enough for them to be one step ahead of host immune cell clonal selection.

Advanced Higher Biology (revised): Antibody techniques

Antibodies are widely used in the detection and identification of specific proteins. Immunoassay techniques use antibodies linked to reporter enzymes to cause a colour change in the presence of a specific antigen. Fluorescent labelling of antibodies in blotting and (immunohistochemical) staining of tissue. Use of monoclonal antibodies in the diagnosis and detection of disease. Use the ELISA technique to identify the presence of specific antigens. To produce stocks of a particular antibody, hybridomas are formed by fusion of a B lymphocyte with a myeloma cell using polyethelene glycol (PEG).

Use of monoclonal antibodies in the diagnosis and detection of disease. Use the ELISA technique to identify the presence of specific antigens.



Advanced Higher Biology: Biotechnology Unit

Protein nature of antibodies produced in response to specific antigens by B lymphocytes. Site and production of B lymphocytes. Preparation of polyclonal sera and its disadvantages. Monoclonals produced from a single B line secreting one specific antibody. Nature of myeloma cells and their hybridisation with lymphocytes using polyethylene glycol (PEG) to produce hybridomas. Use of selective media and screening. Hybridomas only produce one particular monoclonal. Batch culture of secreting hybridomas in fermenters and extraction of pure antibody.

Uses of monoclonal antibodies

Use of monoclonal antibodies in diagnosis and detection of disease. Use of immunoassay (ELISA) techniques involving monoclonal antibodies joined to enzyme; coloured product used to quantify presence of antigen specific to pathogen eg AIDS, meningitis, *Botrytis.* Treatment of disease: tumour-specific antibodies joined to toxins, combine with tumour cells and kill them.



Immune responses

- What triggers immune responses?
 - Signals that 'alert' the innate immune system
- How can we make immune responses to effectively combat many different infections (viruses, bacteria, parasites?)
 - Selective activation of the <u>adaptive</u> immune system
- What controls immune responses once they have started?
 - Without regulation the immune system can cause disease (autoimmunity)

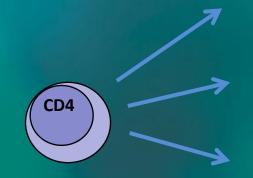


Host immunity

- B lymphocytes make antibodies
- Antibodies work against infections when they are outside cells (neutralising virus infectivity)
- T lymphocytes are important for cell-mediated immunity (CMI)
- CMI works against infections inside cells (can be viruses, bacteria, protozoa). T lymphocytes can recognise and kill infected cells or programme the cells to kill microbes inside them themselves (via cytokines)
- Lymphocyte activation is required how?



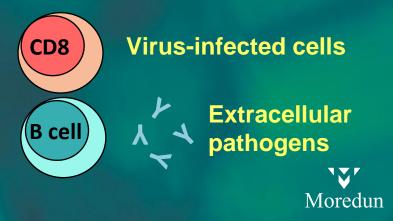
Making the 'right' response



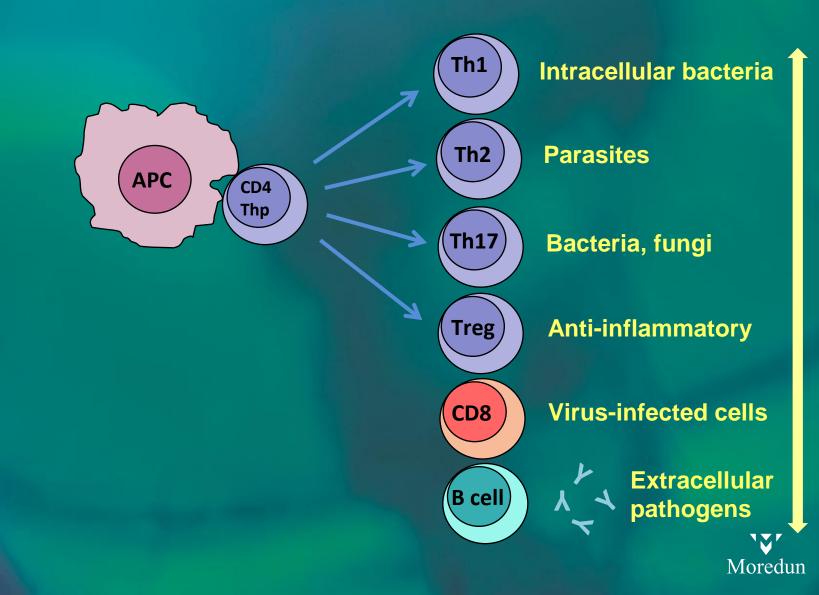
Intracellular bacteria

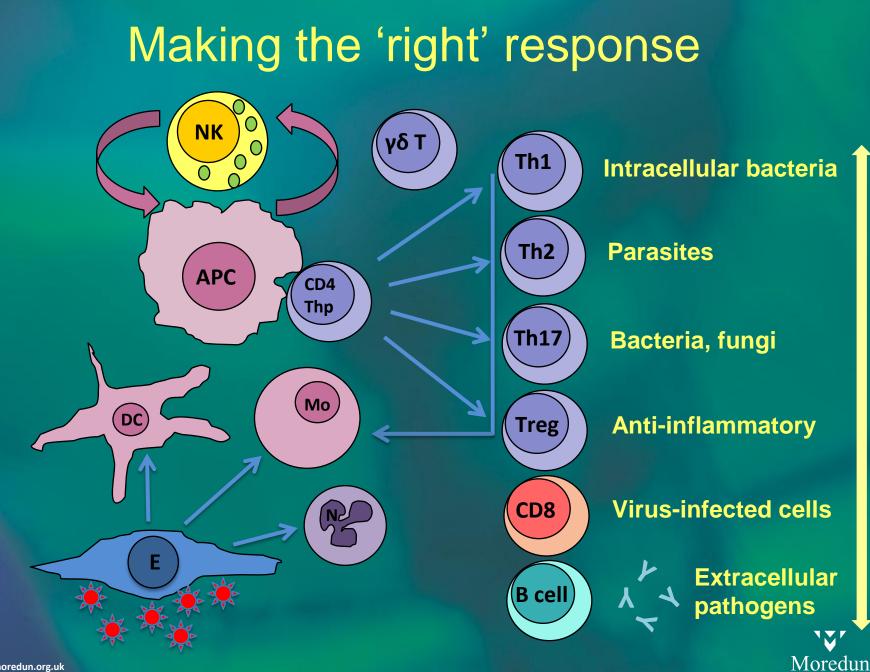
Parasites

Bacteria, fungi



Making the 'right' response





The clonal selection model

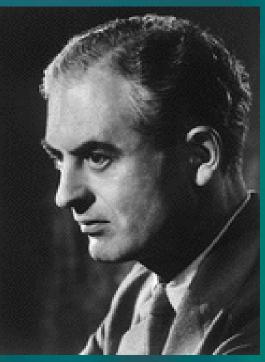


The clonal selection model

Macfarlane Burnet



Peter Medawar





Nobel Prize in Physiology or Medicine 1960 "for the discovery of immunological tolerance"



Clonal selection model

- Lymphocytes express receptors of a single antigenic specificity
- This specificity is genetically determined and precedes antigen encounter
- Antigen only stimulates cells with specific receptors and induces clonal expansion of that lymphocyte population
- Diversity is achieved by random gene rearrangement (*RAG* genes) and somatic mutation



Clonal selection model

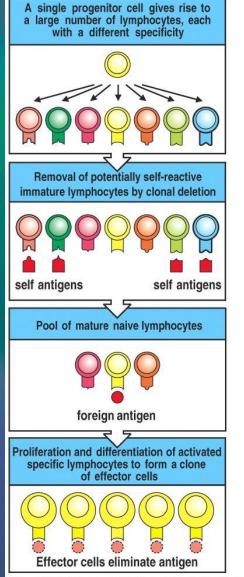
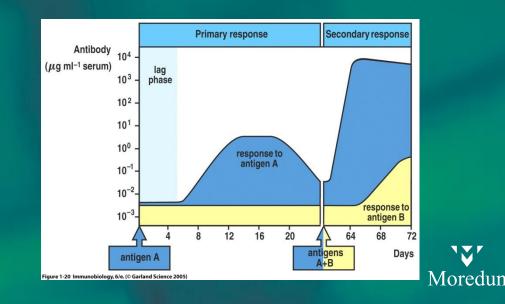
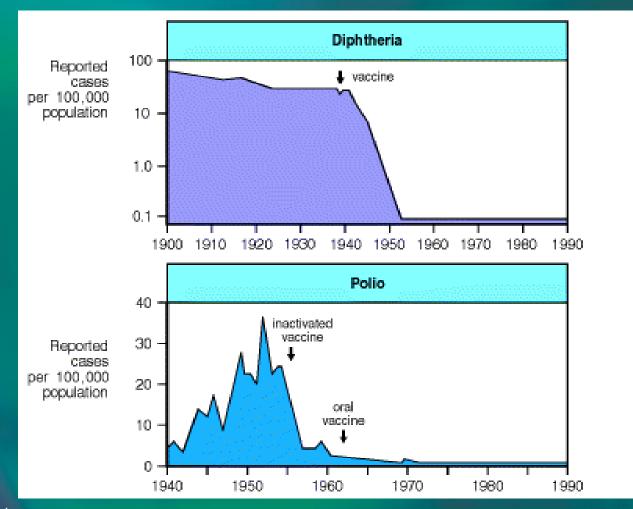


Figure 1-14 Immunobiology, 6/e. (© Garland Science 2005)

This theory has underpinned the formulation of paradigms relating to immunological <u>self tolerance</u> and immunological <u>memory</u> as well as practical applications of <u>vaccine</u> <u>design</u>



Vaccination is a very effective means of preventing infectious diseases



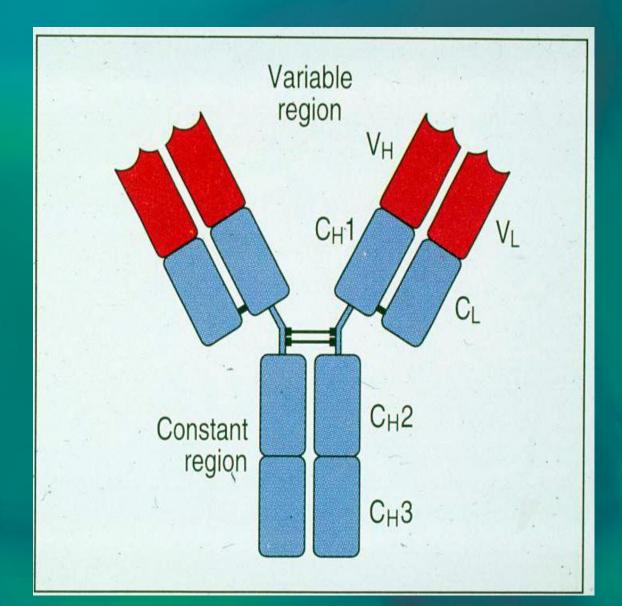


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Production of monoclonal antibodies



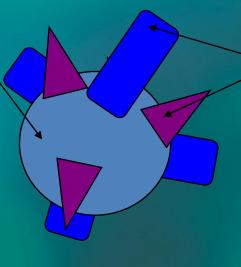
Antibody structure





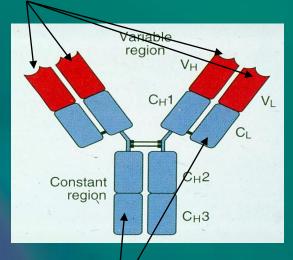
Some technical terms:

Antigen- Any molecule which may be a protein that stimulates antibody production.



Epitope- Unique marker on antigen surface responsible for stimulating antibody production.

Antigen binding sites



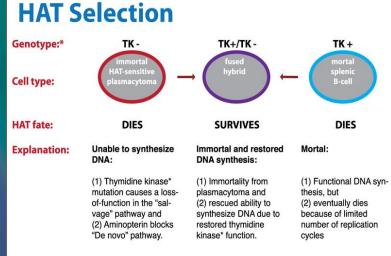
Constant region

Antibody or Immunoglobulin (Ig)- Protein that binds specifically to a particular antigen.
Monoclonal – single epitope specificity
Polyclonal- multiple epitope specificities

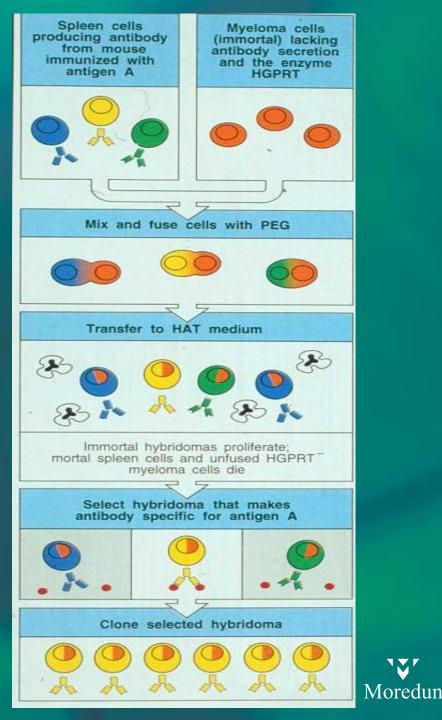




Production of monoclonal antibodies



*HGPRT (hypoxanthine-guanine phosphoribosyltransferase) mutants can be used in place of TK (thymidine kinase) mutants



Use of monoclonal antibodies for therapeutics



Therapeutic monoclonal antibodies

Clinically-approved therapies for cancer, autoimmunity/inflammation, infection, transplant rejection

Exert their effects by:

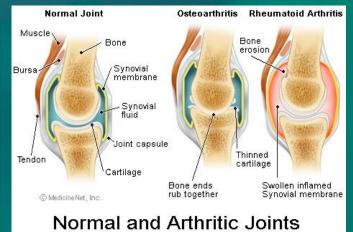
Blocking biological function by neutralising soluble factors or inhibiting cell-cell interactions.

Killing target cells through receptor blockage, activating cytotoxic immune cells or via selective targeting of a cytotoxic compound



Immunotherapy for rheumatoid and psoriatic arthritis

Rheumatoid and psoriatic arthritis are chronic progressive conditions characterised by inflammation in the joints. TNF- α is commonly observed in both conditions.



Conventional therapy relies on non-steroidal antiinflammatory drugs (NSAIDs) and disease-modifying antirheumatic drugs (DMARDs) such as methotrexate. DMARDs are most effective when used early. In patients where DMARDs fail, alternative therapeutics include Infliximab (Remicade®), a chimaeric antibody that neutralises TNF- α . It is an <u>antagonist.</u>

Cancer immunotherapy



Approximately 20-30% of breast cancers over-express the human epidermal growth factor receptor-2 (HER-2) on their cell surface

Trastuzumab is a humanised monoclonal antibody that binds HER-2 and stops cells dividing. It also suppresses angiogenesis (blood vessel development)

Trastuzumab is marketed under the trade name Herceptin®



Use of monoclonal antibodies for diagnostics



ELISA: the definition

Enzyme-Linked ImmunoSorbent Assay...

S S S S S S S S S S Ε F Ε S S S

.....is a test that can measure antibodies from biological fluids such as blood and milk.



Well of serological plate

Why do we use an ELISA test?

 We can use ELISAs to measure antibodies or antigens present in the blood of humans and animals

ELISA process involves multiple steps



ELISA: The works- step 1 of 11

1. Specific antigen is bound to wells of an ELISA plate



Coat with antigen



Representative well of ELISA plate

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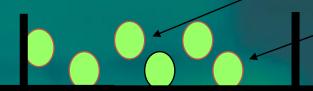
ELISA: step 2 of 11

2. Any excess antigen washed away

Excess antigen washed away

Bound antigen remains





ELISA: step 3 of 11

3. Blocking solution added to ensure the test sample binds correctly

Blocking solution added



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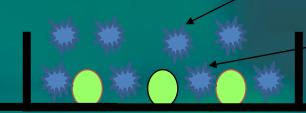
ELISA: step 4 of 11

4. Excess blocking solution is washed away

Excess blocking solution is washed away

Bound block solution remains

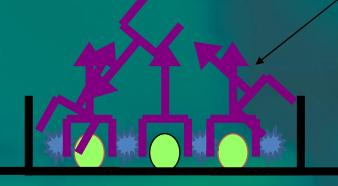




ELISA: step 5 of 11

5. Test sample added to well. Each well is used to test a separate sample

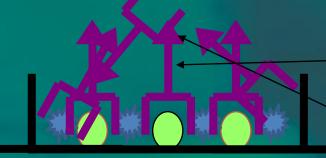
Blood test sample added to well





ELISA: step 6 of 11

6. Specific antibodies bind to antigen, anything else is washed away

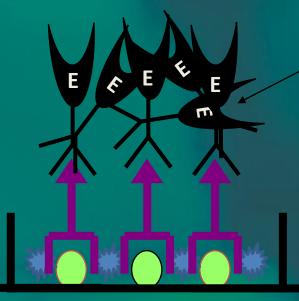


specific antibodies are bound to antigen
Unbound sample washed away



ELISA: step 7 of 11

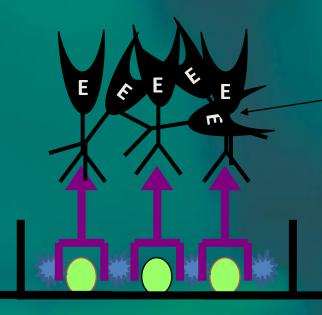
7. Enzyme-coupled detection molecules are added



Enzyme-coupled detection molecules are added



ELISA: step 8 of 11 8. Wash away unbound enzymecoupled detection molecules



Wash away unbound enzymecoupled detection molecules



ELISA: step 9 of 11

S

S

S

S

9. Add colourless substrate. The substrate will react with the enzymes

S

Ε

Ε

S

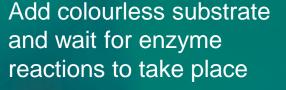
Ε

S

S

S

S





ELISA: step 10 of 11 10. Colourless substrate will undergo a reaction and change colour. The stronger the colour, the more antibody is detected in the well

S

Ε

S

S

Substrate will be

enzyme

converted to a blue

liquid if it binds to the

S

S

S

S

S

S

S

S

S

Ε

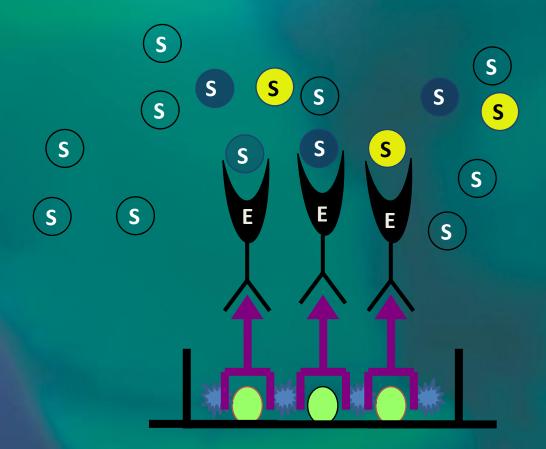
S

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ELISA: step 11 of 11

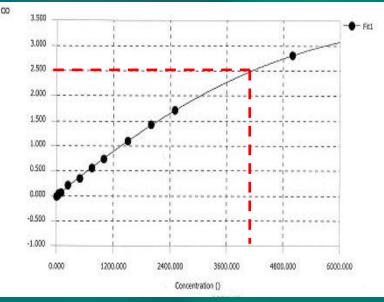
11. The reaction is stopped by adding acid, converting blue substrate solution to yellow





Understanding the test results:

 A spectrophotometer is used to measure intensity of the colourchange. The readings from the machine can be read off the curve to reveal the antibody concentration.





Range of standards to produce curve Kit positive Kit negative

 For example, colour value 2.5
 gives 4200 units of antibody



Advantages of ELISAs as diagnostic tests:

- Only a small sample is needed
- Quick results: usually 2-4hrs
- Simple technique that can be accurately reproduced
- Many samples can be tested at the same time
- Relatively inexpensive

